



Tarım Bilimleri Dergisi  
Tar. Bil. Der.  
Dergi web sayfası:  
www.agri.ankara.edu.tr/dergi

Journal of Agricultural Sciences  
Journal homepage:  
www.agri.ankara.edu.tr/journal

## Genetic Diversity and Population Structure of *Fusarium oxysporum* f. sp. *cepae*, the Causal Agent of Fusarium Basal Plate Rot on Onion, using RAPD Markers

Harun BAYRAKTAR

Ankara University, Faculty of Agriculture, Department of Plant Protection, 06110 Ankara, TURKEY

### ARTICLE INFO

Research Article — Crop Production

Corresponding author: Harun BAYRAKTAR, e-mail: bayrakta@agri.ankara.edu.tr, Tel: +90(312) 596 12 70

Received: 24 May 2010, Received in revised form: 01 November 2010, Accepted: 10 November 2010

### ABSTRACT

Fusarium basal plate rot caused by *Fusarium oxysporum* f. sp. *cepae* is an important disease of onion in many countries of the world. A total of 116 single spore isolates of *Fusarium oxysporum* f. sp. *cepae* representing seven populations from Turkey and one population from Colorado, USA were subjected to molecular marker analysis using Random Amplified Polymorphic DNA (RAPD) technique. Using eleven polymorphic primers, 110 RAPD fragments were obtained with an average of 10 polymorphic bands per primer. Cluster analysis with UPGMA revealed five distinct lineages at arbitrary level of 65% similarity. The majority of *F. oxysporum* f. sp. *cepae* isolates from Turkey were observed to derive probably from the same clonal lineage. Genetic estimates and population differences demonstrated that the isolates from Turkey were significantly distinct from Colorado isolates and that Bursa population was the most divergent among Turkish populations. Cluster analysis of Nei's genetic distances supported populations grouping according to the geographical regions. Comparison of genetic differentiation estimates ( $H_T$ : 0.140,  $H_S$ : 0.124) revealed low levels of genetic differentiation among Turkish populations. Only, 11.4% of total genetic diversity ( $G_{ST}$ ) attributed to differentiation among the geographical populations. Analysis of molecular variance (AMOVA) confirmed that there was low genetic differentiation among populations. The results suggest that RAPD-PCR is a useful method for analyzing genetic variation within and between populations of *F. oxysporum* f. sp. *cepae*.

Keywords: Fusarium basal rot; Genetic diversity; Onion; RAPD-PCR; *Fusarium oxysporum* f. sp. *cepae*

## Soğanda Dip Çürüklüğü Etmeni *Fusarium oxysporum* f. sp. *cepae*' daki Genetik Farklılığının ve Popülasyon Yapısının RAPD Markörleri Kullanılarak İncelenmesi

### ESER BİLGİSİ

Araştırma Makalesi — Bitkisel Üretim

Sorumlu Yazar: Harun BAYRAKTAR, e-posta: bayrakta@agri.ankara.edu.tr, Tel: +90(312) 596 12 70

Geliş tarihi: 24 Mayıs 2010, Düzeltmelerin gelişi: 01 Kasım 2010, Kabul: 10 Kasım 2010

### ÖZET

*Fusarium oxysporum* f. sp. *cepae*' nın sebep olduğu Fusarium dip çürüklüğü, dünyanın birçok ülkesinde soğan bitkisinin önemli bir hastalığıdır. Türkiye'den yedi ve Colorado, ABD'den bir popülasyonu temsil eden toplam 116

*Fusarium oxysporum* f. sp. *cepae* izolatu Random Amplified Polymorphic DNA (RAPD) tekniđi kullanılarak moleküler markör analizi ile incelenmiştir. Kullanılan 11 polimorfik primer ile 110 RAPD fragmenti elde edilmiştir (10 polimorfik bant/primer). Elde edilen verilerin UPGMA ile cluster analizi % 5 benzerlik seviyesinde 5 farklı soy ortaya çıkarmıştır. Türkiye'den elde edilen *Fusarium oxysporum* f. sp. *cepae* izolatlarının büyük çoğunluğunun muhtemelen aynı klonal soydan türediđi gözlemlenmiştir. Genetik deđerlendirmeler ve popülasyon farklılıkları da Türkiye' den elde edilen izolatların Colorado izolatlarından büyük ölçüde farklı olduğunu ve Bursa popülasyonunun Türkiye popülasyonları arasında en farklı popülasyon olduğunu göstermiştir. Nei' nin genetik uzaklığına dayanan cluster analizi ise cođrafik bölgelere göre popülasyon gruplanmasını desteklemiştir. Genetik farklılık deđerlerinin ( $H_T$ : 0.140,  $H_S$ : 0.124) karşılaştırılması ise Türkiye popülasyonları arasında düşük seviyelerde genetik farklılıklar ortaya çıkarmıştır. Toplam genetik farklılığın sadece % 11.4'lük kısmının cođrafik popülasyonlar arasındaki farklılıklardan kaynaklandığı gözlemlenmiştir. Moleküler varyans analizi de (AMOVA) popülasyonlar arasındaki düşük genetik farklılıkları doğrulamıştır. Bu sonuçlar RAPD-PCR tekniđinin *Fusarium oxysporum* f. sp. *cepae* popülasyonları arasındaki ve içerisindeki genetik varyasyonun deđerlendirilmesinde oldukça faydalı olduğunu göstermiştir.

Anahtar sözcükler: *Fusarium* dip çürüklüğü; Genetik farklılık; Soğan; RAPD-PCR; *Fusarium oxysporum* f. sp. *cepae*

© Ankara Üniversitesi Ziraat Fakültesi

## 1. Introduction

Onion (*Allium cepa* L.) is a crop of economic importance and widely grown in many countries. Turkey is a major onion producing country with a cultivated area of 95,000 ha and a production of 2 175,343 t yr<sup>-1</sup> as dry and green onion (Faostat 2010). *Fusarium oxysporum* f. sp. *cepae* W.C Snyder & H.N Hansen, the causal agent of Fusarium basal rot is a common disease in onion growing regions worldwide, causing economically significant losses in both field and storage (Abawi & Lorbeer 1971; Sumner 1995). The pathogen infects the root and basal plate areas of onion plants. The disease symptoms include curving, wilting, yellowing, and eventually dying back of the leaves from the tips. In addition to onion, the pathogen can cause yield losses on other *Allium* species such as garlic, chive and shallot (Sumner 1995). Although *F. oxysporum* f. sp. *cepae* has a high degree of pathogenic variability on onion plants, the classification of different races has not been described yet (Sumner 1995; Özer et al 2004).

The most effective and economical strategy to control soil borne pathogens is the use of resistant cultivars. Some intermediate and long day onion cultivars have proved to be resistant to Fusarium basal rot while resistant short day cultivars are not developed. Also, several studies have been performed regarding the genetic structure of onion resistance, but have not exactly clarified the

resistance mechanism to the pathogen (Cramer 2000).

Genetic analysis of plant pathogen populations is fundamental to the improving of breeding programmes and disease management methods to control the pathogens. Different methods have been used to assess genetic variation within populations of *F. oxysporum* f. sp. *cepae* such as Vegetative Compatibility Grouping (VCGs), Amplified Fragment Length Polymorphisms (AFLPs), Restriction Fragment Length Polymorphism (RFLP) analysis of Internal Transcribed Spacer (ITS) and Intergenic Spacer (IGS) regions of rDNA (Swift et al 2002, Galván et al 2008; Dissanayake et al 2009, Bayraktar et al 2010). These studies have revealed that *F. oxysporum* f. sp. *cepae*, which includes different genetic lineages is a polyphyletic group. RAPD technique is a powerful genetic approach for population studies in fungi because it is simple, not expensive and easy to use. This method has been successfully used to resolve genetic variability within or between formae speciales of *F. oxysporum* (Assigbetse et al 1994; Bentley et al 1994; Cramer et al 2003). However, RAPD markers have not been used to detect phylogenetic groups within populations of this pathogen. Also, little information is known about the genetic complexity of *F. oxysporum* f. sp. *cepae* populations in Turkey and there is a gap for future studies to identify resistance sources to this pathogen.

The objective of the present study was to determine the geographical relationships and the extent of genetic diversity among isolates of *F. oxysporum* f. sp. *cepae* using random amplified polymorphic DNA markers.

## 2. Materials and Methods

### 2.1. Fungal isolates

One hundred isolates were recovered from infected onion bulbs collected from Bursa, Ankara, Eskişehir, Yozgat, Çorum, Tokat and Amasya provinces located in three regions of Turkey during 2007 surveys (Table 1 & Figure 1). The isolates were purified by single spore isolation and grown on potato dextrose agar (PDA) plates at 25°C in a 12 h dark/light cycle. Fungal cultures were preserved on filter papers at 8°C. Morphological identification was carried out according to Nelson et al (1983) and Leslie & Summerell (2006). Also, the identification was confirmed by PCR using species-specific FOF/R 1 primers, producing a single fragment of 340-bp (Mishra et al 2003). The pathogenicity of isolates was determined by seedling and bulb inoculation methods used in a previous study (Bayraktar et al 2010). Pathogenic variability among the isolates was found to vary from 1 to 5, most of them were highly virulent on onion plants. Also, the 16 Colorado isolates assigned into five VCG groups (Swift et al 2002) and provided by H. F. Schwartz at Colorado State University were included in this study (Table 1).

### 2.2. DNA extraction

Genomic DNA for PCR was extracted using a modified method of Reader & Broda (1985). Cultures were grown in 100 ml of potato dextrose broth medium on an orbital shaker (150 rpm) at 25°C for 7 days. Mycelia from cultures were collected on Miracloth by filtration and ground in liquid nitrogen. The powdered samples of mycelium were suspended in 500 µl of extraction buffer (200 mM Tris-HCl pH 8.5, 25 mM NaCl, 25 mM EDTA, 0.5% SDS) and incubated for 30 min at 65°C. After incubation, equal volume of phenol-chloroform-isoamylalcohol (25:24:1 v/v/v) was added into the mixture and centrifuged at 13 000 x g for 15 min. The supernatant was incubated with

10 µl RNase A (10 mg ml<sup>-1</sup>) to remove RNA at 37°C for 1 h. The suspension was re-extracted with chloroform-isoamylalcohol and centrifuged as above. The DNA was precipitated with isopropanol (1:1 v/v), rinsed with ethanol, suspended in TE buffer (10 mM Tris-HCl, 1mM EDTA, pH 8.0) and stored at -20°C until use.

### 2.3. Amplification conditions

RAPD analysis was performed in a total volume of 25 µl reaction containing 10x PCR buffer, 1.5 mM of MgCl<sub>2</sub>, 0.32 µM of primer, 0.125 mM of dNTPs and 0.6 unit of *Taq* DNA polymerase (MBI, Fermentas). Amplifications were carried out in a thermal cycler (Whatman-Biometra Model T1, Goettingen, Germany) programmed as follows: 40 cycles of 94°C 20 sec, 36°C 1 min, 72°C 1 min and followed by an extension step of 8 min 72°C. Thirty eight primers selected from different sets (Operon Technologies Inc. Alameda, CA.) were evaluated for their ability to produce polymorphic bands on a subset of 15 isolates.

The PCR products were separated electrophoretically at 100 V on 1.4% agarose gels in 1xTAE (40 mM Tris-acetate and 1.0 mM EDTA) buffer (Sambrook et al 1989). The gels were stained with ethidium bromide and visualized by Gene Tools bio-imaging system from SynGene software (Cambridge, England). All experiments were repeated twice.

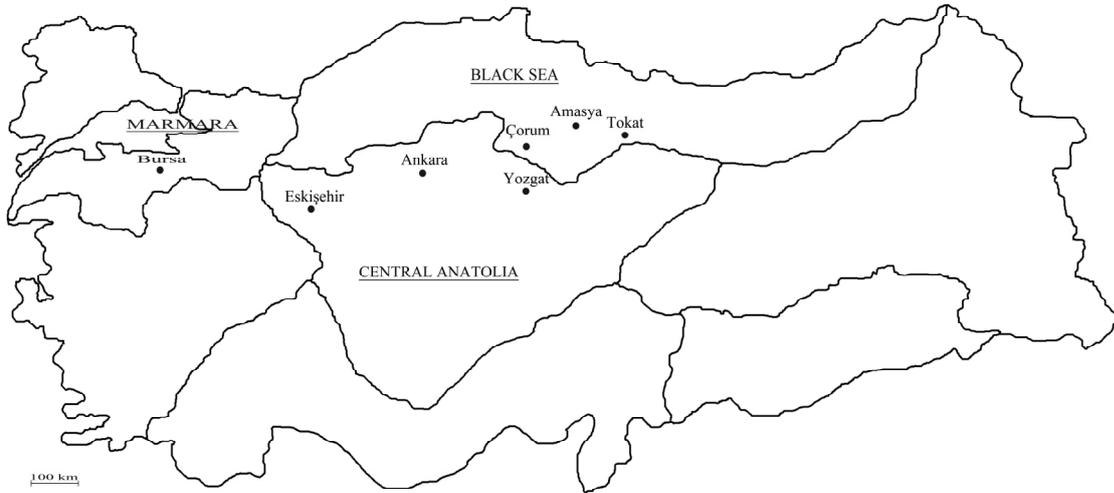
### 2.4. Data analysis

The presence or absence of a given RAPD fragment was determined for each isolate. All scored bands were assembled in a rectangular data matrix. The genetic similarity matrix was created with Jaccard's coefficient of similarity and subjected to cluster analysis with an unweighted pair-grouped method with arithmetic average (UPGMA) to generate a dendrogram using NTSYS-pc numerical taxonomy package, ver. 2.0 (Rohlf 1998). Genetic diversity and population differentiation parameters were calculated using Popgene software ver. 1.32 (Yeh et al 1999). The estimates of Nei's (1973) gene diversity (*h*), Shannon's information index (*I*), polymorphic loci (%), genetic distance and genetic identity, total gene diversity (*H<sub>T</sub>*), gene diversity within populations (*H<sub>S</sub>*), genetic differentiation

**Table 1-Geographical origin and the number of *Fusarium oxysporum* f. sp. *cepae* isolates used in this study**  
Çizelge 1-Çalışmada kullanılan *Fusarium oxysporum* f. sp. *cepae* izolatlarının coğrafik orjinleri ve sayısı

Regions	Populations	Isolate code	Total
Marmara	Bursa	s1, s7, s9, s13, s18, s20, s21, s23, s24, s25, s26, s27, s75, s79, s81, s82, s86, s87, s88, s103, s105	21
	Çorum	s2, s5, s6 (VCG 0423), s8, s10, s28, s29, s31, s32, s38, s74, s80, s93, s96, s101	15
Black Sea	Amasya	s17, s19, s45, s48, s49, s51, s52, s53, s54, s56, s57, s58, s59, s60, s62, s63, s66, s69, s72, s73, s85, s92, s94, s97	24
	Tokat	s4, s12, s14, s37, s30, s84, s90, s102	8
Central Anatolia	Ankara	s64, s65, s67, s68, s70, s76, s77, s78, s104, s106	10
	Eskişehir	s3, s15, s16, s41, s42, s43, s46, s47, s91, s98, s100	11
	Yozgat	s33, s34, s35, s36, s39, s40, s44, s71, s83, s89, s99	11
Colorado <sup>a</sup>		VCG	
		22, 25, 27, 28, 201a, 201d, 204a	0421
		31, 32	0422
		41, 42, 47, 48, 50	0423
		21	4
	23	5	

<sup>a</sup> Isolates from Colorado (Swift et al 2002) were provided by H. F. Schwartz at Colorado State University

**Figure 1-Map showing locations of seven populations of *Fusarium oxysporum* f. sp. *cepae* sampled from Turkey**

Şekil 1-Türkiye' den örneklenen *Fusarium oxysporum* f. sp. *cepae*' nin yedi popülasyonunun yerlerini belirten harita

( $G_{ST}$ ), and gene flow ( $N_M$ ) were calculated among populations of *F. oxysporum* f. sp. *cepae*. UPGMA cluster analysis of Nei's (1972) genetic distances was generated to represent the relationships among

eight populations. Analysis of molecular variance (AMOVA) between and within populations was performed by the program Arlequin ver. 3.0 (Excoffier et al 2005) using 1023 permutations.

### 3. Results

#### 3.1. Genetic diversity

Among 38 primers tested on a subset of 15 isolates, the eleven primers that gave reproducible and scorable polymorphisms were used for molecular analysis of all isolates (Table 2). These primers produced a total of 110 reproducible fragments among the 116 isolates of *F. oxysporum* f. sp. *cepae*, all of them were polymorphic, giving a ratio of 10 polymorphic bands/primer. Among the eleven primers, the most informative primer was OPA-3, producing 16 polymorphic bands while primer OPE-12 was the least informative primer with 6 fragments. The size of RAPD fragments ranged from 400 to 4.230 bp.

RAPD analysis of genomic DNA from the pathogenic isolates revealed the presence of five lineages at the arbitrary level of 65% similarity (Figure 2). Lineage A consisted of two isolates, 22 (VCG 0421) and 23 from Colorado. Lineage B included all isolates representing VCG 0422 and VCG 0423 as well as isolate s20. Eight isolates from five provinces of Turkey clustered into lineage C together with VCG 0421 isolates and isolate 21 from Colorado. Lineage D was the largest cluster occupying 64.66% of total samples. Isolate s58 was the most divergent in this lineage. Lineage E consisted of 15 isolates from different provinces. However, there was no correlation between clustering in dendrogram, pathogenic variability and geographical origin of the tested isolates. The cophenetic correlation coefficient between Jaccard's similarity matrices and the dendrogram of RAPD data was  $r=0.98$ , showing very good fit.

#### 3.2. Population structure of *F. oxysporum* f. sp. *cepae*

The polymorphic loci in each population ranged from 20.91% to 84.55% with an average of 51.36% (Table 3). Colorado population showed the highest percentage of polymorphic loci among all populations while Eskişehir population exhibited the lowest amount of polymorphism. Similarly, it was observed that the estimates of Turkish populations were much lower than Colorado population. The estimates of gene diversity ( $h$ ) for all loci in each of the Turkish populations ranged

from 0.056 (Eskişehir) to 0.199 (Bursa). The value of Colorado population was 0.346. The gene diversity index calculated for all populations was 0.21. Similarly, the values of Shannon information index ( $I$ ) were similar to Nei's (1973) gene diversity index for all geographical populations, ranging from 0.089 to 0.498 among all populations.

The total gene diversity ( $H_T$ ) was  $0.195 \pm 0.016$  for all populations of *F. oxysporum* f. sp. *cepae*. The mean within-population gene diversity ( $H_S$ ) was  $0.152 \pm 0.012$ , which accounted for 77.95% of the total genetic diversity (Table 3). The proportion of the total genetic diversity attributable to the population differentiation ( $G_{ST}$ ) was 0.223 over all loci. When population structure was analyzed by excluding the Colorado population, the mean total and within gene diversities ( $H_T$ ,  $H_S$ ) were  $0.140 \pm 0.020$  and  $0.124 \pm 0.015$  among all Turkish populations. Only, 11.4% of the total gene diversity ( $G_{ST}$ ) was attributed to differences among populations.

Genetic analysis showed that the highest genetic identity (0.993) occurred between populations Eskişehir and Yozgat while the lowest genetic identity (0.799) was recorded between populations Eskişehir and Colorado (Table 4). The UPGMA dendrogram based on Nei's genetic distances among populations is shown in Figure 3. Colorado population clustered separately from Turkish populations in dendrogram while all Turkish populations sampled from the same geographical regions clustered together in dendrogram. The populations of Çorum, Amasya and Tokat located in Black Sea region were clustered to each other at the level of 0.013 genetic distance while Bursa population was closely related to three populations of Black Sea region, but formed a distinct subgroup with genetic distance of 0.019. The populations of Ankara, Eskişehir and Yozgat located in Central Anatolia region were very closely related and formed a different cluster of dendrogram with a low level (0.010) of genetic distance.

Results of AMOVA gave a nearly identical pattern of population differentiation; all RAPD variation was distributed among isolates (95.19%) within populations (Table 5). Gene flow ( $N_M$ ) was 3.895 among Turkish populations.

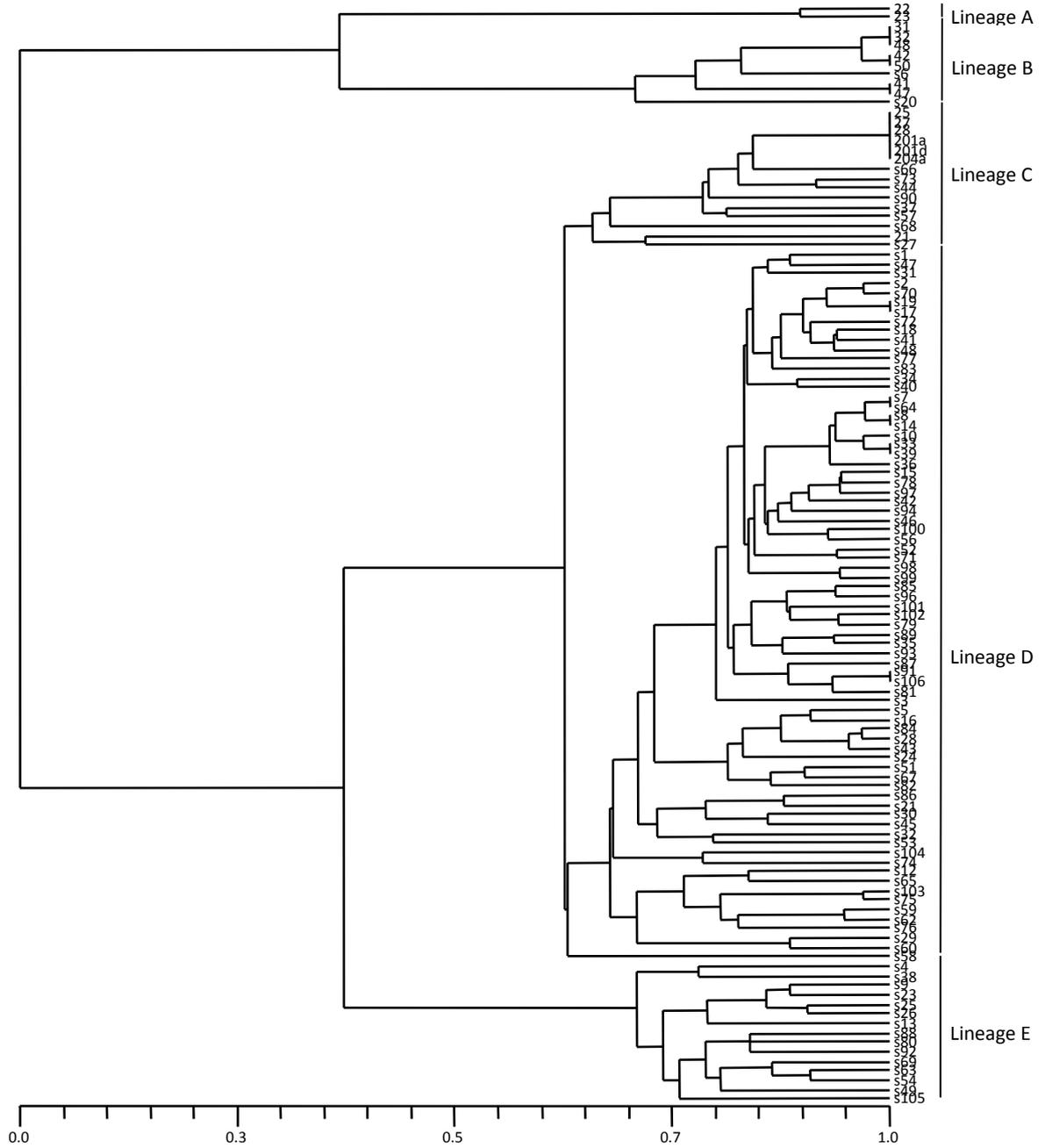
**Table 2-Primers used and the number and size of bands obtained from *Fusarium oxysporum* f. sp. *cepae* isolates**Çizelge 2-Kullanılan primerler ve *Fusarium oxysporum* f. sp. *cepae* izolatlarından elde edilen bantların sayısı ve büyüklüğü

Primer	Sequence 5'-3'	Total band number	Size bp min-max
OPA-03	AGTCAGCCAC	16	400-2.900
OPA-04	AATCGGGCTG	11	490-2.650
OPB-18	CCACAGCAGT	13	400-2.390
OPE-01	CCCAAGGTCC	9	490-4.230
OPE-12	TTATCGCCCC	6	450-1.860
OPF-10	GGAAGCTTGG	8	550-2.400
OPF-12	ACGGTACCAG	12	480-3.700
OPH-02	TCGGACGTGA	9	540-2.200
OPH-05	AGTCGTCCCC	7	650-2.310
OPI-09	TGGAGAGCAG	9	430-2.560
OPK-12	TGGCCCTCAC	10	560-3.980

**Table 3-Genetic diversity estimates within and among eight populations of *Fusarium oxysporum* f. sp. *cepae***Çizelge 3-*Fusarium oxysporum* f. sp. *cepae*'nin sekiz popülasyonu arasındaki ve içindeki genetik farklılık tahminleri

Populations	N	Polymorphic loci, %	h	I
Bursa	21	77.27	0.199±0.184	0.312±0.251
Çorum	15	79.09	0.174±0.142	0.289±0.203
Amasya	24	51.82	0.143±0.172	0.224±0.252
Tokat	8	42.73	0.138±0.177	0.212±0.261
Ankara	10	30.91	0.095±0.160	0.147±0.236
Eskişehir	11	20.91	0.056±0.122	0.089±0.186
Yozgat	11	23.64	0.065±0.134	0.104±0.201
Colorado	16	84.55	0.346±0.195	0.498±0.260
Turkish populations			H <sub>T</sub> : 0.140±0.020 H <sub>S</sub> : 0.124±0.015	G <sub>ST</sub> : 0.114
All populations			H <sub>T</sub> : 0.195±0.016 H <sub>S</sub> : 0.152±0.012	G <sub>ST</sub> : 0.223

N: Number of isolates h: Nei's (1973) gene diversity I: Shannon's Information index (Lewontin 1972). H<sub>T</sub>: total gene diversity; H<sub>S</sub>: gene diversity within populations; and G<sub>ST</sub>: the proportion of total gene diversity found among populations

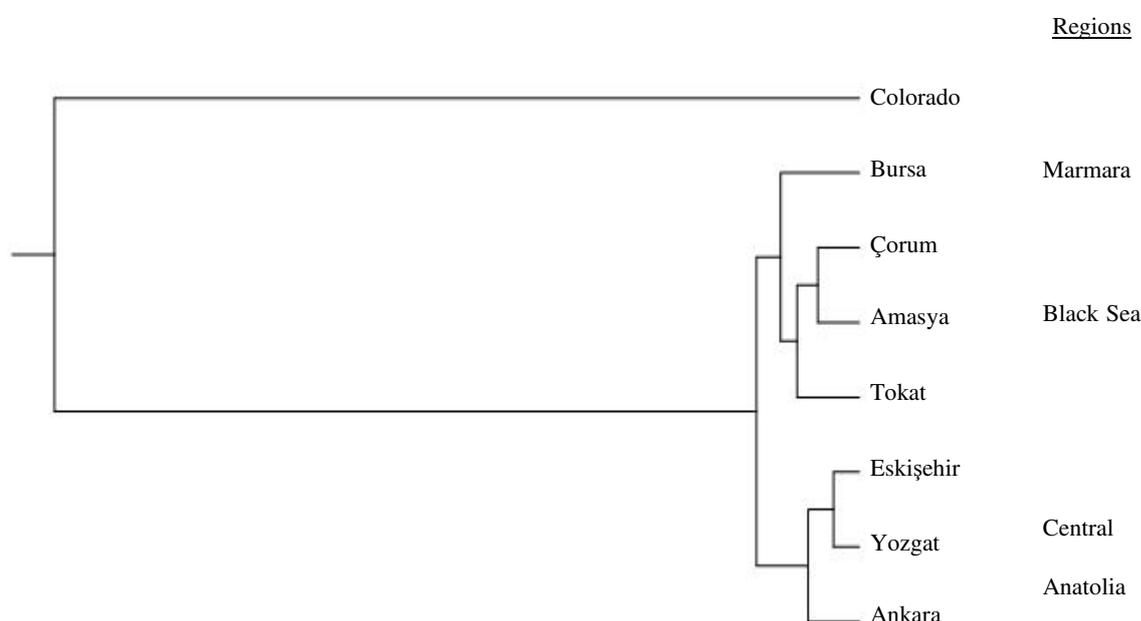


**Figure 2-UPGMA dendrogram constructed for 116 isolates of *Fusarium oxysporum* f. sp. *cepae* sampled from Turkey and Colorado based on Jaccard's coefficient of similarity**

*Şekil 2-Türkiye ve Colorado' dan örneklenen Fusarium oxysporum f. sp. cepae' nın 116 izolatu için Jaccard' ın benzerlik katsayısı ile oluşturulan UPGMA dendogramı*

**Table 4-Genetic identity between pairwise combinations of *Fusarium oxysporum* f. sp. *cepae* populations**  
*Çizelge 4-Fusarium oxysporum* f. sp. *cepae*'nin populasyon çiftleri arasındaki genetik benzerlik

Populations	Bursa	Çorum	Amasya	Tokat	Ankara	Eskişehir	Yozgat
Çorum	0.983						
Amasya	0.985	0.989					
Tokat	0.974	0.981	0.988				
Ankara	0.967	0.987	0.986	0.981			
Eskişehir	0.952	0.984	0.979	0.972	0.987		
Yozgat	0.954	0.983	0.980	0.973	0.988	0.993	
Colorado	0.841	0.846	0.821	0.824	0.814	0.799	0.806

**Figure 3-Dendrogram based on Nei's genetic distance showing relationships among the eight geographic populations of *Fusarium oxysporum* f. sp. *cepae***

*Şekil 3-Fusarium oxysporum* f. sp. *cepae*'nin sekiz coğrafik popülasyonu arasındaki ilişkiyi göstermek için Nei' nin genetik uzaklığı ile oluşturulan dendrogram

**Table 5-The AMOVA results for seven populations of *Fusarium oxysporum* f. sp. *cepae* from Turkey**  
*Çizelge 5-Fusarium oxysporum* f. sp. *cepae*'nin yedi Türkiye popülasyonu için belirlenen AMOVA sonuçları

Source of variation	Degree of freedom	Sum of squares	Variance components	Percentage of variation	P value
Among populations	6	80.175	0.396	4.810	
Within populations	93	729.595	7.845	95.190	<0.008
Total	99	809.770	8.241		

P value, probability of obtaining equal or larger value determined by 1.023 permutations

#### 4. Discussion

In this study, genetic diversity within and among eight *F. oxysporum* f. sp. *cepae* populations sampled from Turkey and Colorado, USA was evaluated using RAPD markers. The RAPD procedure proved to be a useful tool for assessing genetic variability within this pathogen. Earlier to this, a high level of VCG diversity was observed among Turkish isolates, all isolates constituted unique VCGs except for one isolate. Also, a broad pathogenic and genetic diversity was detected among isolates of *F. oxysporum* f. sp. *cepae*. Restriction digests on IGS region of rDNA classified the isolates from Turkey and Colorado into 13 groups (Bayraktar et al 2010).

The results obtained in this study indicated a high degree of genetic variability within populations of *F. oxysporum* f. sp. *cepae*. All isolates clustered into five lineages in the dendrogram. A higher level of genetic diversity was observed among the isolates from Colorado than Turkish isolates. The sixteen isolates representing VCGs from 5 groups classified into three lineages related to VCG groups. However, no differences were detected among clustering in dendrogram of VCG 0422 and 0423 isolates, indicating that they were very closely related. VCG 0421 isolates clustered into two lineages, A and C. The Colorado isolates 21, 23, not assigned into VCG groups showed a high level of genetic similarity with VCG 0421 isolates, indicating that these isolates probably derived from the same lineages. The similar genetic relationships among RAPD analysis and VCGs were observed on populations of *F. oxysporum* from different hosts such as banana (Bentley et al 1994), common bean (Woo et al 1996), cucumber (Vakalounakis & Fragkiadakis 1999), and strawberry (Nagarajan et al 2006). Due to high level of VCG diversity among Turkish isolates, RAPD markers provided more satisfactory results in the evaluation of genetic diversity among the isolates. The isolates from seven provinces of Turkey grouped into four lineages. RAPD analysis proved that the majority of Turkish isolates were genetically homogeneous, and distinct from Colorado isolates. Ten isolates from different provinces shared the same lineages with Colorado

isolates, which may result from the introduction of different genetic groups to onion production areas in Turkey with the transportation of infected seeds and bulbs. Lineages D and E included isolates from Turkey only. However, RAPD analysis did not reveal a relationship between genetic and pathogenic variability of Turkish isolates. Similarly, Dissanayake et al (2009) separated *F. oxysporum* isolates obtained from Welsh onion into two genetic groups with 61.6% similarity, but not detected clear-cut relationship between clustering in the PCR-RFLP dendrogram and pathogenicity test. These results were in agreement with the previous studies which fail to provide a relationship between RAPD data and pathogenic groups of *Fusarium oxysporum* (Hernandez et al 1999; Freeman & Maymon 2000; Cramer et al 2003).

RAPD analysis revealed similar polymorphic patterns among most of the isolates from different provinces of Turkey, indicating that *F. oxysporum* f. sp. *cepae* has a homogeneous population structure across the country. For example, isolates s7 (Bursa) and s14 (Tokat) collected from 780 km apart from each other included in the same lineage. However, no relationship was found between genetic variability and geographical origin of the isolates. Similar results were obtained by Galván et al (2008) who classified 43 isolates of *F. oxysporum* f. sp. *cepae* representing various geographical regions into two groups based on AFLP markers from three primer combinations. They observed high genetic similarity among the isolates representing different countries, but not detected correlation between AFLP groups and the origins of isolates. The previous studies failed to show a positive correlation between geographical origin and DNA polymorphisms of *Fusarium* spp. (Migheli et al 1998; Gargouri et al 2003).

The estimates of population structure revealed a low, but significant level of genetic variability in each of Turkish populations. The similar levels of gene diversity were detected in the populations sampled from the same regions of Turkey. The genetic diversity was almost negligible especially in Ankara, Eskişehir, and Yozgat populations located in Central Anatolia region. The values of polymorphic loci and Shannon's information indicated similar levels of genetic variability in

Turkish populations. In contrast to Turkish populations, high degrees of gene and genotype diversity were detected in Colorado population. The results of Swift et al (2002) supported high level of genetic diversity in the Colorado population which was placed into five VCGs.

The estimates of Nei's (1972) genetic distance and identity between the populations provided a better understanding of genetic relationships. UPGMA analysis of genetic distances among the populations confirmed that the Turkish populations were genetically distinct from the Colorado population, and high degree of genetic similarity was present among Turkish populations separated by geographical distances. Bursa population is geographically closer to the populations of Central Anatolia, although genetically more similar to the populations of Black Sea, indicating that genetic variability among populations of *F. oxysporum* f. sp. *cepae* has developed regardless of geographical distance in Turkey. The low  $G_{ST}$  and AMOVA values revealed little evidence for geographical subdivision among seven populations. Similarly, low levels of genetic differentiation among geographic populations and high genetic variability have been reported on asexually reproducing fungi such as *F. oxysporum* f. sp. *vasinfectum* (Wang et al 2006), *F. culmorum* (Gargouri et al 2003) and *Aspergillus niger* (Pekarek et al 2006). The values of gene flow were adequate to prevent genetic differentiation among populations of *F. oxysporum* f. sp. *cepae*.

## 5. Conclusion

These results indicated that RAPD markers, revealing five clonal lineages among isolates of *F. oxysporum* f. sp. *cepae* sampled from eight geographical populations were well-suited for the clarification of genetic diversity within and among the pathogen populations. The estimates of genetic structure revealed that the populations from Turkey and Colorado were genetically different, and genetic differentiation among Turkish populations was low. The knowledge of population structure among geographical populations of *F. oxysporum* f. sp. *cepae* will be useful in breeding programmes for developing resistant onion cultivars to the pathogen

as well as providing more insights into the molecular evolution of this formae speciales.

## Acknowledgements

This research was supported by Ankara University (Scientific Research Project). I am thankful to Prof. H. F. Schwartz (Department of Bioagricultural Sciences and Pest Management, Colorado State University, USA) for Colorado isolates.

## References

- Abawi G S & Lorbeer J W (1971). Reaction of selected onion varieties to infection by *Fusarium oxysporum* f. sp. *cepae*. *Plant Disease Reporter* **55**: 1000-1004
- Assigbetse K B, Fernandez D, Dubois M P & Geiger J P (1994). Differentiation of *Fusarium oxysporum* f. sp. *vasinfectum* races on cotton by Random Amplified Polymorphic DNA (RAPD) analysis. *Phytopathology* **84**: 622-626
- Bayraktar H, Türkkän M & Dolar F S (2010). Characterization of *Fusarium oxysporum* f. sp. *cepae* from onion in Turkey based on vegetative compatibility and rDNA RFLP analysis. *Journal of Phytopathology* **158**: 691-697
- Bentley S, Pegg K G & Dale J L (1994). Optimization of RAPD-fingerprinting to analyze genetic variation within populations of *Fusarium oxysporum* f. sp. *cubense*. *Journal of Phytopathology* **142**: 64-78
- Cramer C S (2000). Breeding and genetics of Fusarium basal rot resistance in onion. *Euphytica* **115**: 159-166
- Cramer R A, Byrne P F, Brick M A, Panella L, Wickliffe E & Schwartz H F (2003). Characterization of *Fusarium oxysporum* isolates from common bean and sugar beet using pathogenicity assays and random-amplified polymorphic DNA markers. *Journal of Phytopathology* **151**: 352-360
- Dissanayake M L M, Kashima R, Tanaka S & Ito S I (2009). Pathogenic variation and molecular characterization of *Fusarium* species isolated from wilted Welsh onion in Japan. *Journal of General Plant Pathology* **75**: 37-45
- Excoffier L, Laval G & Schneider S (2005). Arlequin ver. 3.0: An Integrated Software Package for Population Genetics Data Analysis: University of Berne Switzerland
- Faostat (2010). Statistical database. Available: <http://faostat.fao.org>
- Freeman S & Maymon M (2000). Reliable detection of

- the fungal *Fusarium oxysporum* f. sp. *albedinis*, causal agent of bayoud disease of date palm, using molecular techniques. *Phytoparasitica* **28**(4): 341-348
- Galván G A, Koning-Boucoiran C F S, Kopman W J M, Burger-Meijer K, González P H, Waalwijk C, Kik C & Scholten O E (2008). Genetic variation among *Fusarium* isolates from onion and resistance to *Fusarium* basal rot in related *Allium* species. *European Journal of Plant Pathology* **121**: 499-512
- Gargouri S, Bernier L, Hajlaoui M R & Marrakchi M (2003). Genetic variability and population structure of the wheat foot rot fungus, *Fusarium culmorum*, in Tunisia. *European Journal of Plant Pathology* **109**: 807-815
- Hernandez J F, Posada M A & Arbelaez G (1999). Identification of molecular markers of *Fusarium oxysporum* f. sp. *dianthi* by RAPD. *Proceedings of the International Symposium on cut flowers in the Tropics*, pp 123-131
- Leslie J F & Summerell B A (2006). *The Fusarium laboratory manual*. Blackwell, Ames, pp 388.
- Lewontin R C (1972). The apportionment of human diversity. *Evolutionary Biology* **6**: 381-398
- Migheli Q, Briatore E & Garibaldi A (1998). Use of random amplified polymorphic DNA (RAPD) to identify races 1, 2, 4 and 8 of *Fusarium oxysporum* f. sp. *dianthi* in Italy. *European Journal of Plant Pathology* **104**: 49-57
- Mishra P K, Fox R T V & Culham A (2003). Development of a PCR-based assay for rapid and reliable identification of pathogenic Fusaria. *FEMS Microbiology Letters* **218**: 329-332
- Nagarajan G, Kang S W, Nam M H, Song J Y, Yoo S J & Kim H G (2006). Characterization of *Fusarium oxysporum* f. sp. *fragariae* based on vegetative compatibility group, random amplified polymorphic DNA and pathogenicity. *The Plant Pathology Journal* **22**(3): 222-229
- Nei M (1972). Genetic distance between populations. *American Naturalist* **106**: 283-292
- Nei M (1973). Analysis of gene diversity in subdivided populations. *Proceedings of the National Academy of Sciences USA* **70**: 3321-3323
- Nelson P E, Toussoun T A & Marasas W F O (1983). *Fusarium Species. An illustrated manual for identification*. Pennsylvania State University Press, pp 193
- Özer N, Koycu D, Chilosi D & Magro P (2004). Resistance to *Fusarium* basal rot of onion in greenhouse and field and associated expression of antifungal compounds. *Phytoparasitica* **32**: 388-394
- Pekarek E, Jacobson K & Donovan A (2006). High levels of genetic variation exist in *Aspergillus niger* populations infecting *welwitschia mirabilis* hook. *Journal of Heredity* **97**(3): 270-278
- Reader U & Broda P (1985). Rapid preparation of DNA from filamentous fungi. *Letters in Applied Microbiology* **1**: 17-20
- Rohlf F I (1998). NTSYS-pc. Numerical taxonomy and multivariate analysis system, version 2.0. Applied Biostatistics, New York, USA
- Sambrook J, Fritsch E F & Maniatis T (1989). *Molecular cloning: A laboratory manual*, 2<sup>nd</sup> edn. Cold Spring Harbor Laboratory Press, New York, pp 1659
- Sumner D R (1995). *Fusarium* basal plate rot, In: Schwartz H.F. and Mohan S.K. (eds.). *Compendium of Onion and Garlic Diseases*. St. Paul, MN, USA, APS press, pp 10-11
- Swift C E, Wickliffe E R & Schwartz H F (2002). Vegetative compatibility groups of *Fusarium oxysporum* f. sp. *cepae* from onion in Colorado. *Plant Disease* **86**: 606-610
- Vakalounakis D J & Fragkiadakis G A (1999). Genetic diversity of *Fusarium oxysporum* isolates from cucumber: differentiation by pathogenicity, vegetative compatibility, and RAPD fingerprinting. *Phytopathology* **89**: 161-168
- Wang B, Brubaker C L, Tate W, Woods M J, Matheson B A & Burdon J J (2006). Genetic variation and population structure of *Fusarium oxysporum* f. sp. *vasinfectum* in Australia. *Plant Pathology* **55**: 746-755
- Woo S L, Zoina A, Del Sorbo G, Lorito M, Nanni B, Scala F & Noviello C (1996). Characterization of *Fusarium oxysporum* f. sp. *phaseoli* by pathogenic races, VCGs, RFLPs, and RAPD. *Phytopathology* **86**: 966-973
- Yeh F C, Yang R C, Boyle T J, Ye Z H & Mao J X (1999). Popgene ver. 1.32, the user-friendly shareware for population genetic analysis. Molecular Biology and Biotechnology Centre, University of Alberta, Edmonton, Canada